



## **Generation of a gene-corrected isogenic control cell line from an Alzheimer's disease patient iPSC line carrying a A79V mutation in PSEN1**

Pires, Carlota; Schmid, Benjamin; Petræus, Carina; Poon, Anna Fong-Yee; Nimsanor, Natakarn; Nielsen, Troels Tolstrup; Waldemar, Gunhild; Hjermand, Lena Elisabeth; Nielsen, Jørgen Erik; Hyttel, Poul; Freude, Kristine

*Published in:*  
Stem Cell Research

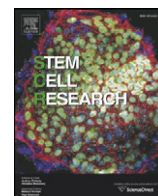
*DOI:*  
[10.1016/j.scr.2016.08.002](https://doi.org/10.1016/j.scr.2016.08.002)

*Publication date:*  
2016

*Document version*  
Publisher's PDF, also known as Version of record

*Document license:*  
[CC BY-NC-ND](#)

*Citation for published version (APA):*  
Pires, C., Schmid, B., Petræus, C., Poon, A. F-Y., Nimsanor, N., Nielsen, T. T., Waldemar, G., Hjermand, L. E., Nielsen, J. E., Hyttel, P., & Freude, K. (2016). Generation of a gene-corrected isogenic control cell line from an Alzheimer's disease patient iPSC line carrying a A79V mutation in *PSEN1*. *Stem Cell Research*, 17(2), 285-288. <https://doi.org/10.1016/j.scr.2016.08.002>



Lab Resource: Stem Cell Line

# Generation of a gene-corrected isogenic control cell line from an Alzheimer's disease patient iPSC line carrying a A79V mutation in *PSEN1*



Carlota Pires<sup>a</sup>, Benjamin Schmid<sup>b</sup>, Carina Peträus<sup>a</sup>, Anna Poon<sup>a</sup>, Natakarn Nimsanor<sup>c</sup>, Troels T. Nielsen<sup>d</sup>, Gunhild Waldemar<sup>d</sup>, Lena E. Hjermand<sup>d</sup>, Jørgen E. Nielsen<sup>d</sup>, Poul Hyttel<sup>a</sup>, Kristine K. Freude<sup>a,\*</sup>

<sup>a</sup> Faculty of Health and Medical Sciences, University of Copenhagen, Grønnegaardsvej 7, 1870 Frederiksberg C, Denmark

<sup>b</sup> Bioneer A/S, Hoersholm, Denmark

<sup>c</sup> Faculty of Medical Technology, Mahidol University, Bangkok, Thailand

<sup>d</sup> Danish Dementia Research Centre, Rigshospitalet, University of Copenhagen, Blegdamsvej 9, 2100 Copenhagen Ø, Denmark

## ARTICLE INFO

## Article history:

Received 15 July 2016

Accepted 3 August 2016

Available online 7 August 2016

## ABSTRACT

Alzheimer's disease (AD) is a progressive and irreversible neurodegenerative disease causing neural cell degeneration and brain atrophy and is considered to be the most common form of dementia. We previously generated an induced pluripotent stem cell (iPSC) line from an AD patient carrying an A79V mutation in *PSEN1* as an *in vitro* disease model. Here we generated a gene-corrected version from this hiPSC line by substituting the point mutation with the wild-type sequence. The reported A79V-GC-iPSCs line is a very useful resource in combination with the A79V-iPSC line in order to study pathological cellular phenotypes related to this particular mutation.

© 2016 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Resource table

Name of stem cell construct	A79V-GC-hiPSC
Institution	University of Copenhagen
Person who created resource	Carlota Pires, Kristine K Freude
Contact person and email	Kristine K Freude: <a href="mailto:kkf@sund.ku.dk">kkf@sund.ku.dk</a>
Date archived/stock date	February 2016
Origin	Human induced pluripotent cell line (hiPSCs)
Type of resource	Biological reagent: gene-corrected induced pluripotent stem cells, originally derived from skin fibroblasts of a <i>PSEN1</i> A79V heterozygous mutation patient
Sub-type	Induced pluripotent stem cells (iPSCs)
Key transcription factors	<i>hOCT4</i> , <i>hSOX2</i> , <i>hKLF4</i> , <i>hL-MYC</i> , <i>hLIN28</i> , and shRNA against TP53 (Addgene plasmids 27077, 27078 and 27080; Okita et al., 2011)
Authentication	Identity and purity of cell line confirmed by mutation sequencing, karyotyping and pluripotency markers (Fig. 1)
Link to related literature (direct URL links and full references)	<a href="http://dx.doi.org/10.1016/j.scr.2016.01.002">http://dx.doi.org/10.1016/j.scr.2016.01.002</a> Li T, Pires C, Nielsen TT, Waldemar G, Hjermand LE, Nielsen JE, Dinnyes A, Hyttel P, Freude KK. Generation of induced pluripotent stem cells (iPSCs) from an Alzheimer's disease patient carrying an A79V mutation in <i>PSEN1</i> , Stem Cell Research. 2016; 16: 229–232
Information in public databases	N/A

## Resource details

The study was approved by the “De Videnskabssetiske Komiteer for Region Hovedstaden” (protocol number H-4-2011-157), Copenhagen, Denmark and written informed consent was obtained in all cases. To protect the patient family privacy, no personal patient information is presented here. Mutations in presenilin 1 (*PSEN1*) gene are the most common known causes of inherited Alzheimer's disease (AD).

A human induced pluripotent stem cell (hiPSC) line was previously generated from a skin biopsy obtained from a 48-year-old woman carrying a heterozygous mutation in exon 4 of the *PSEN1* gene, which causes a change in amino acid A79V. Episomal plasmids carrying gene sequences for *hOCT4*, *hSOX2*, *hKLF4*, *hL-MYC*, *hLIN28* and a short hairpin against TP53 (Okita et al., 2011) were used to reprogram the fibroblasts into iPSCs, successfully establishing integration and feeder-free iPSCs. This cell line has been described previously as a *bona fide* iPSC line with normal karyotype (Li et al., 2016).

The A79V-hiPSC (c.236 C > T) was gene-corrected with the CRISPR/Cas9 system (Ran et al., 2013) by replacing the point mutation “T” with the wild-type nucleotide “C” (Fig. 1A). The nucleotide substitution/correction was validated by sequencing (Fig. 1B). Gene-integrity was also checked by confirming that the DNA sequence surrounding the mutation and the CRISPR cutting site were not altered, with no frameshifts or other mutation(s) being generated upon gene editing, confirming successful gene correction of A79V-hiPSC, named A79V-GC-hiPSC (“GC” = gene corrected). The sequencing of the isogenic line confirmed the c.236 T > C replacement in exon 4 of the *PSEN1*

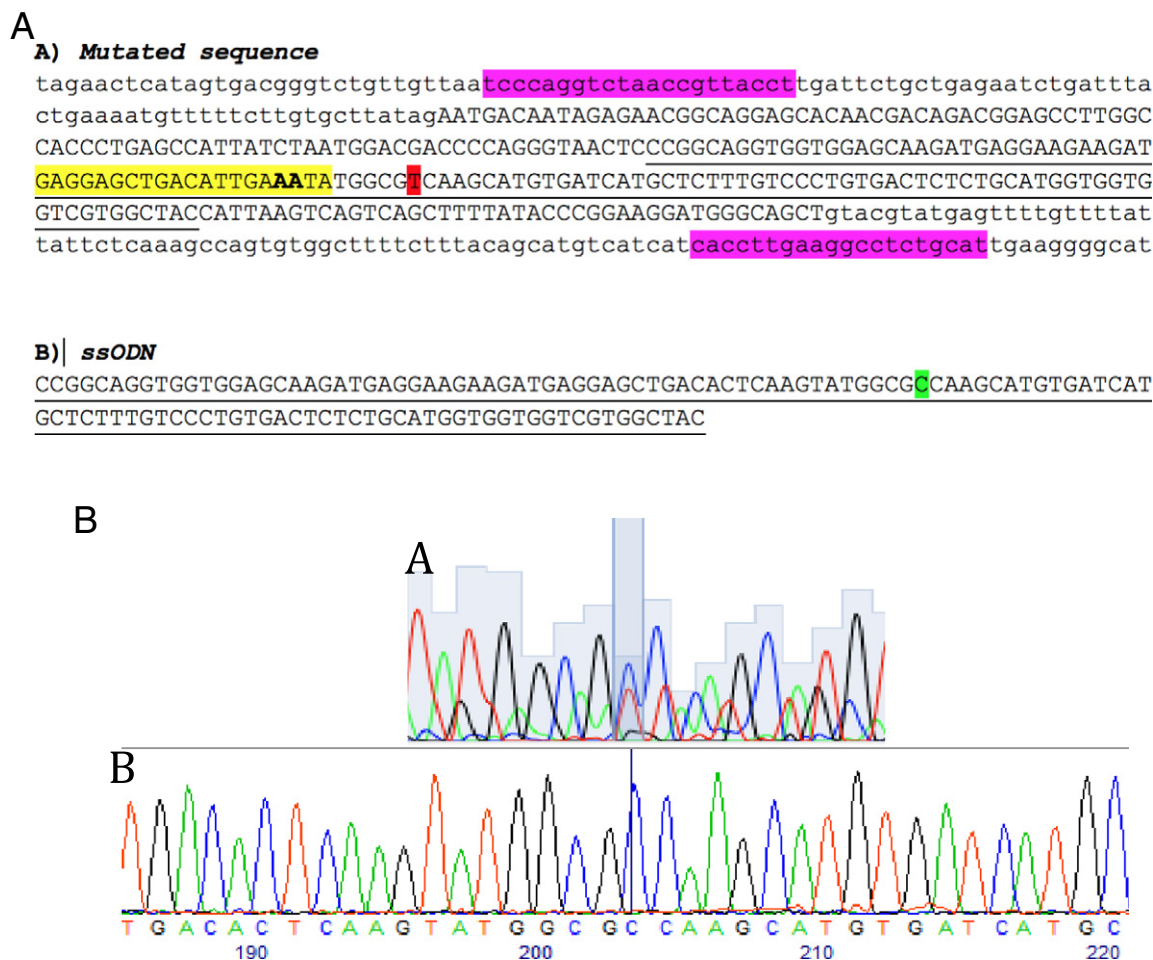
DOI of original article: <http://dx.doi.org/10.1016/j.scr.2016.01.002>.

\* Corresponding author.

E-mail address: [kkf@sund.ku.dk](mailto:kkf@sund.ku.dk) (K.K. Freude).

<http://dx.doi.org/10.1016/j.scr.2016.08.002>

1873-5061/© 2016 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).



**Fig. 1.** A. CRISPR/Cas9 and ssODN used to repair the point mutation in A79V-hiPSC. A) Genomic sequence surrounding the mutation site: mutated nucleotide (T, red); sgRNA recognition site containing 20 bp (yellow); CRISPR cutting site between the 17th and 18th bp (bold); forward and reverse primers (pink). B) ssODN with 120 bp, 60 bp upstream and 60 bp downstream the mutation site containing the WT nucleotide (C, green). B. Sequencing of exon 4 of the *PSEN1* gene in hiPSCs. A) Heterozygous c.236C > T substitution in the mother line previously published. B) Successful correction of the point mutation (WT nucleotide – C; Mutated nucleotide – T). C. Immunocytochemical detection in hiPSCs of the pluripotency markers OCT4, NANOG, SSEA3, SSEA4, TRA1-60 and TRA1-81. Scale bars correspond to 200  $\mu$ m. D. Representative karyotype of A79V-GC-iPSCs.

gene corresponding to the wild-type (healthy) sequence (Fig. 1B). Immunocytochemical (ICC) analysis of the pluripotency markers OCT4, NANOG, SSEA3, SSEA4, TRA-1-60 and TRA-1-81 (Fig. 1C) confirmed the gene-edited line maintained its pluripotency like the mother line previously published. Finally, karyotyping was performed to confirm that no chromosomal aberrations were introduced in the isogenic line upon gene correction (Fig. 1D).

## Materials and methods

### CRISPR design

Isogenic gene-corrected controls were obtained using the CRISPR/Cas9 technology in combination with single-stranded oligonucleotides (ssODNs), which provided the wild-type template for gene-correction.

CRISPRs were designed with the help of the software available at <http://crispr.mit.edu/>. The single-guide RNAs (sgRNA) were ordered from Sigma and the cloning of CRISPRs followed the protocol from Ran et al. (2013), with a single plasmid containing both the sgRNA and the Cas9 (pSpCas9(BB)-2A-Puro (PX459) V2.0; addgene ID 48139).

ssODNs were designed to contain 120 bp in total, with 60 bp upstream and 60 bp downstream of the cutting site. Screening of clones was performed with restriction digestions where the wild-type sequence was recognized (and cut) by specific restriction enzymes (EheI, 5'-GGC↓GCC-3', ThermoScientific), while the mutated sequence

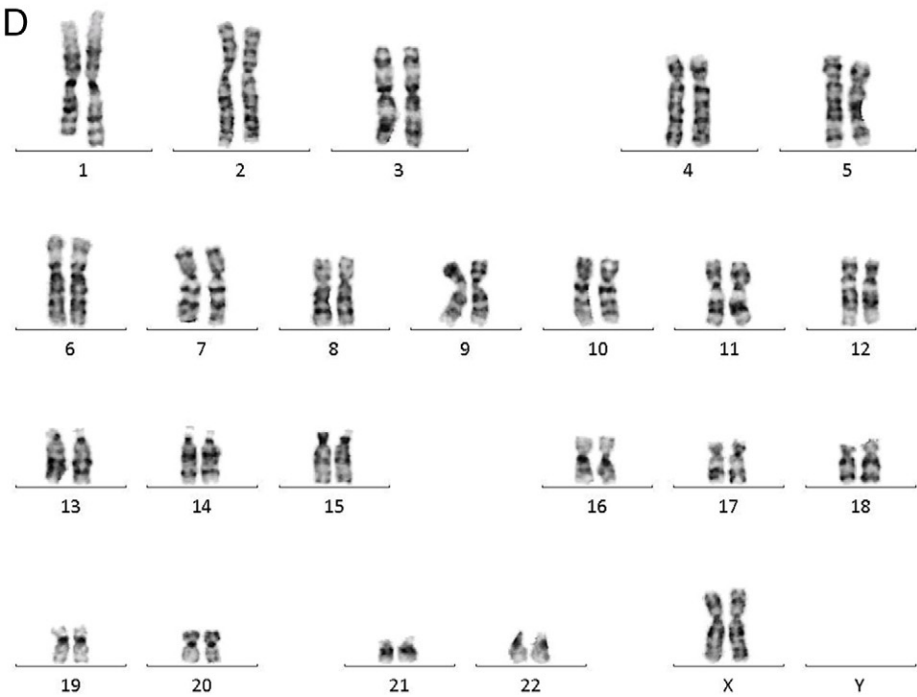
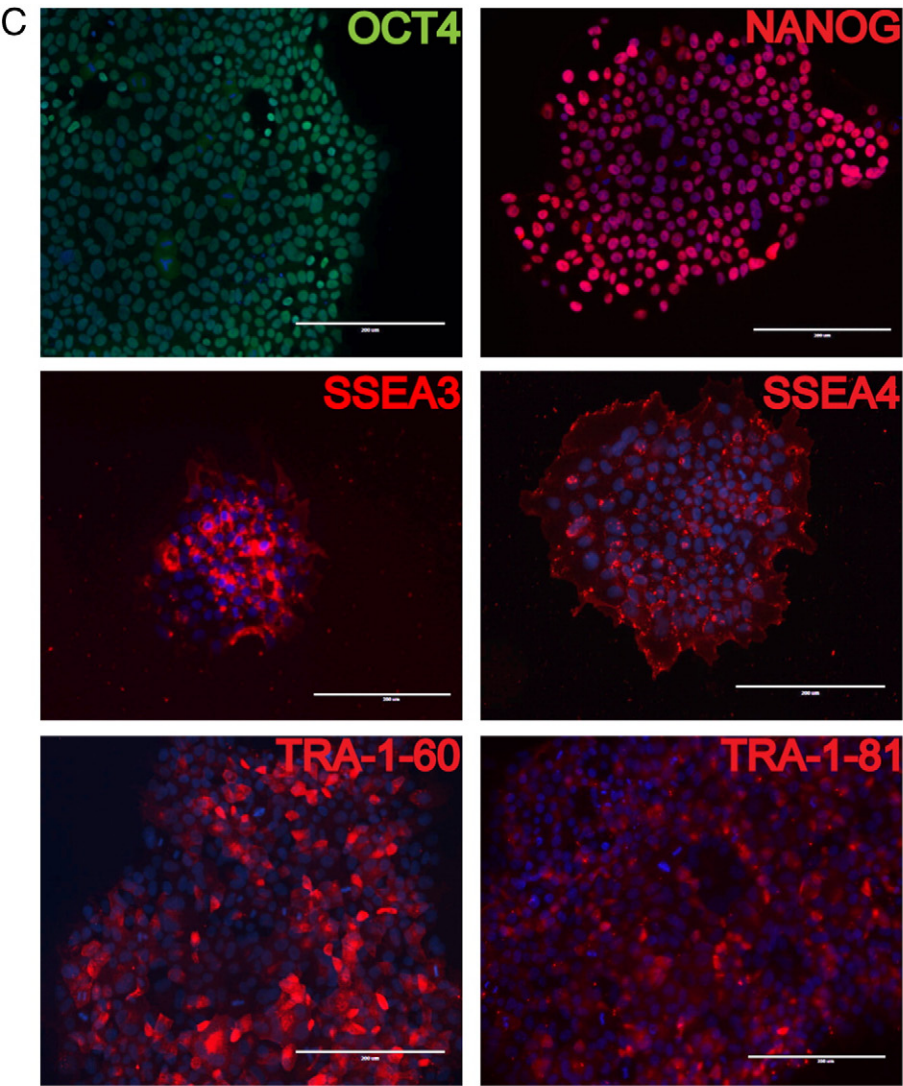
was not, which means EheI only digested clones where the ssODN was successfully integrated.

### Nucleofection

A79V-hiPSCs growing on matrigel-coated (Corning Bioscience) dishes with E8 medium (Gibco) were detached with Accutase (Gibco).  $2 \times 10^6$  cells were nucleofected with 10  $\mu$ g of CRISPR/Cas9 plasmid and 100 pmol of ssODN using the Amaxa 4D Nucleofector (program CA137, Lonza) and P3 Primary Cell Kit (Lonza) according to the manufacturer's instructions. Nucleofected hiPSCs were plated back in a matrigel-coated dish with E8 medium supplemented with 1 mM ROCK inhibitor (Sigma). hiPSCs were subjected to puromycin selection 4 h after nucleofection and allowed to recover for 4–5 days. Surviving (resistant) colonies were manually picked into 96 well plates coated with matrigel with E8 and then expanded for genotyping and sequencing.

### Genotyping and restriction assays

DNA for genotyping was extracted using the prepGem™ Kit (ZyGEM, PT10500) (Qiagen). PCR genotyping was performed using TEMPase Hot Start DNA Polymerase (Ampliqon) according to the manufacturer's instructions, using an annealing temperature of 60 °C. The following primers were designed to cover the mutation and





sgRNA cutting site, generating a PCR product of 409 base pairs: A79V\_Forward2 5'-TCCCAGGTCTAACCGTTACCT-3' and A79V\_Reverse2 5'-ATGCAGAGGCCTTCAAGGTG-3'. PCR products were digested using the restriction enzyme EheI (ThermoScientific) according to the manufacturer's instructions and clones showing correct digestion patterns were then sequenced to confirm the correction and that no frame-shifts or other mutations were introduced.

### Sequencing

Sanger sequencing of exon 4 of *PSEN1* gene was carried out in an ABI PRISM 310 Genetic Analyzer using a designed set of primers that flank the mutation site (A79V\_Forward2 5'-TCCCAGGTCTAACCGTTACCT-3' and A79V\_Reverse2 5'-ATGCAGAGGCCTTCAAGGTG-3') according to standard procedures.

### Immunofluorescence staining

hiPSCs were fixed with 4% paraformaldehyde (PFA) in PBS for 15 min and stained by standard immunofluorescence staining procedures. The primary antibodies (Table 1) were visualized with the secondary antibodies Alexa488 or Alexa594 diluted 1:200 (Life technologies) and counterstained with Hoechst bisbenzimidazole 33,258. Images were acquired with a Leica DMRB-fluorescence microscope (Leica Microsystems, Wetzlar, Germany).

### Karyotyping

A79V-GC-hiPSCs were treated for 45 min with KaryoMAX colcemid (Life Technologies), harvested in fresh fixative consisting of 25% acetic acid and 75% methanol and sent for G-band karyotyping (Institute of Medical Genetics and Applied Genomics, University of Tübingen, Tübingen, Germany).

### Verification and authentication

Karyotyping was performed by Institute of Medical Genetics and Applied Genomics, University of Tübingen (Tübingen, Germany) and a

**Table 1**

Primary antibodies used for immunocytochemistry.

	Antibody and host species	Dilution	Company and catalog number
Pluripotency	Rabbit anti-NANOG	1:500	Peptrotech, 500-P236
	Goat anti-OCT4	1:500	Santa Cruz, sc-8628
	Rat anti-SSEA3	1:100	Biolegend, 330302
	Mouse anti-SSEA4	1:100	Biolegend, 330402
	Mouse anti-Tra-1-60	1:200	Biolegend, 330602
	Mouse anti-Tra-1-81	1:200	Biolegend, 330702

minimum of 20 metaphases were analyzed. The results showed a normal 46, XX karyotype, without any detectable abnormalities (Fig. 1D). A79V-GC-hiPSC line identity and purity was confirmed by sequencing of *PSEN1* (Fig. 1B) and ICC for pluripotency genes expression (Fig. 1C).

### Acknowledgements

We would like to thank Ulla Poulsen and Ida Jørring for excellent technical assistance. We thank the following agencies for financial support: the European Union 7th Framework Program (PIAP-GA-2012-324451-STEMMAD), Innovation Fund Denmark "Brainstem" (4108-00008B). Carlota Pires is further supported by the Programme of Excellence 2016 (Copenhagen as the next leader in precise genetic engineering CDO2016: 2016CDO04210) from the University of Copenhagen.

### References

- Li, T., Pires, C., Nielsen, T.T., Waldemar, G., Hjermand, L.E., Nielsen, J.E., Dinnyes, A., Hyttel, P., Freude, K.K., 2016. Generation of induced pluripotent stem cells (iPSCs) from an Alzheimer's disease patient carrying an A79V mutation in *PSEN1*. *Stem Cell Res.* 16, 229–232.
- Okita, K., Matsumura, Y., Sato, Y., Okada, A., Morizane, A., Okamoto, S., Hong, H., Nakagawa, M., Tanabe, K., Tezuka, K., Shibata, T., Kunisada, T., Takahashi, M., Takahashi, J., Saji, H., Yamanaka, S., 2011. A more efficient method to generate integration-free human iPS cells. *Nat. Methods* 8, 409–412.
- Ran, F.A., Hsu, P.D., Wright, J., Agarwala, V., Scott, D.A., Zhang, F., 2013. Genome engineering using the CRISPR-Cas9 system. *Nat. Protoc.* 8, 2281–2308.